



Comparative Study of Bioremediation of Organophosphate Pesticide Contaminated Soil using Landfarming Technique in Bauchi Metropolis.

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Abstract

Excessive and continuous dispersion of pesticide which are toxic heterogeneous compound in the environment results in environmental pollution with ecological effects (like increase in death rate of humans and animals) that requires remediation either by stimulating the indigenous microorganism with introduction of nutrients (biostimulation) or through the inoculation of an enriched mixed microbial culture into the soil (bioaugmentation). To demonstrate the potential use of bioremediation in soil contaminated with organophosphate pesticide (OPP), a laboratory study with the objective of evaluating and comparing the effects of bioattenuation, biostimulation, bioaugmentation and hybrid of bioaugmentation and biostimulation was performed. Each wooden Microcosm (A1-A4) contained 5% (w/w) of OPP artificially contaminated in the soil as a sole source of carbon and energy. After 10weeks of remediation, the results revealed that hybrid of bioaugmentation and biostimulation (A4) has the highest organophosphate pesticide removal rate (87.5%) followed by A3 (67.2%), A2 (59.4%), and the lowest organophosphate pesticide A1 (42.0%). Total Heterotrophic Bacteria Count revealed that bioremediation actually took place; A4 had the highest maximum bacteria growth ($3.41E+05CFU/g$) at the 6th week, followed by A3 ($2.96 E+05CFU/g$), A2 ($2.85 E+05CFU/g$), and A1 ($1.71E+05CFU/g$).

Keywords: Bioremediation, Biostimulation, Bioaugmentation, Organophosphate Pesticide, wooden microcosms.

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I. Introduction

Pesticide is widely used in Nigeria; there have been an increase in the usage of pesticide since its introduction in the early fifties for cocoa production. It has been estimated that about 125,000 - 130,000 metric tons of pesticides are applied every year in Nigeria (Asogwa and Dongo, 2009; Agarry *et al.*, 2013). Pesticide therefore can be defined as any chemical substance or mixture of substances intended for preventing, destroying, repelling, or mitigating the effect of any pest of plants and animals. They include herbicides, insecticides, rodenticides, fungicides, molluscides, nematocides, avicides, repellents and attractants used in agriculture, public health, horticulture, food storage or a chemical substance used for a similar purpose (REF). There are currently 140 organophosphate compounds being used as pesticides and as plant growth regulators around the world (kang *et al.*, 2006; Agarry *et al.*, 2013). In addition, it has been reported to be the world's largest selling herbicide in 2013, and Chinese manufacturers are the world's largest producers of glyphosate and its precursors (Steinrucken and Amrhein, 1980; Schonbrunn *et al.*, 2001; Stephen *et al.*, 2008; Mink *et al.*, 2012; Changpeng *et al.*, 2015). The United States Environmental Protection Agency and the European Union consider that there is no potential for the herbicide glyphosate to pose a health risk to humans. Furthermore, early epidemiological studies did not find associations between long term low level exposure to glyphosate and any disease (Aparico *et al.*, 2013; Changpeng *et al.*, 2015), but with the heavy use of fungicides, insecticides and herbicides such as the glyphosate formulation Roundup in agriculture, residues in soils are a growing problem (Zhang *et al.*, 2012;

Piel *et al.*, 2012; He *et al.*, 2014; Li *et al.*, 2014; Changpeng *et al.*, 2015). Indeed, a field test showed that lettuce, carrots, and barley contained glyphosate residues up to one year after the soil was treated with 4.15 kg of glyphosate per hectare (Albers *et al.*, 2009; Yamada *et al.*, 2009; Changpeng *et al.*, 2015). Glyphosate is readily degraded by soil microbes to carbon dioxide and amino methyl phosphonic acid (AMPA) that adsorbs strongly to soil and is not expected to move vertically below the top six inch soil layer. In view of compelling evidence of health effects on humans based on studies especially in developed countries and weak implementation of government policy on regulation / ban or surveillance program for pesticides levels in the environment and foods in Nigeria, there is need for evaluation of organophosphate pesticides detoxification. The objective of this study is to examine, evaluate, and compare the methods of natural attenuation, biostimulation, bioaugmentation, and Hybrid biostimulation and bioaugmentation in the bioremediation of soil contaminated with organophosphate pesticide.

II. Materials and Methods

Soil Sample Collection

An un-impacted soil samples with little or no history of pre and post treatment of pesticide from Agricultural and Bio-resources Engineering Farms (Soil and Water Field), Abubakar Tafawa Balewa University (ATBU), Bauchi State were collected from the surface layer of the vadose zone 15 to 30 cm below the land surface. The soil samples were air-dried, homogenized, passed through a 2 mm (pore size) sieve to enhance proper mixing and extract consisting mainly of stones and dead plant debris discarded, the sample was then stored in a polyethylene bag at room temperature of 29⁰C. The sieved soil was contaminated artificially with Organophosphate Pesticide to a pollutant level of 50,000ppm and its moisture content level was increased from 1.37 to 15 using distilled water. The soil matrix was properly mixed at ambient temperature (25-30⁰C). Each of the contaminated soil (5kg) was stacked into four wooden boxes lined with polyethylene bags internally to prevent the leaching. Each box had dimension of 10cm height X 30cm length X 30cm width with soil layer 1.27cm deep. Various treatment options were prepared according to Table 1.

Table 1: Composition of Various Treatment Options.

Microcosm	Sample	Sample Mass(g)	Treatment
1	A ₁	5000	Sample + No Heat + H ₂ O
2	A ₂	5000	Sample + No Heat + Fortified Cow Dung+ H ₂ O
3	A ₃	5000	Sample + Heat + S ₁ &S ₂ + H ₂ O
4	A ₄	5000	Sample + Heat + S ₁ &S ₂ + Fortified Cow Dung + H ₂ O

Key Note:

S₁= *Bacillus subtilis*

S₂= *Pseudomonas aeruginosa*

Wooden microcosm under Organic manure treatment had a C: N: P ratio of 100:10:1

Experimental Design for the Bioremediation of Glyphosate Spiked Soil

Four wooden boxes used as bioreactors were prepared for each treatment as shown in Figure1, designated as bioattenuation (treatment A₁), biostimulation (treatment A₂), bioaugmentation (treatment A₃) and combined biostimulation and bioaugmentation (treatment A₄). Each bioreactor contained 5 kg of soil contaminated and well mixed with 100 ml of 50 g of Organophosphate pesticide (5% w/v) dissolved in water. The bioreactor under treatment A₂ and A₄ was amended with 8.0 kg of organic manure (cow dung)(biostimulation) and combined biostimulation and bioaugmentation. The bioreactor under treatment (A₃ and A₄) was amended with 50 ml each of inoculum (2.6×10⁵CFU/g combine *Bacillus subtilis* & *Pseudomonas aeruginosa*) (bioaugmentation), respectively. The bioreactor under treatment A₁ was not amended with either Organic manure fertilizer and/or inoculum (natural bioattenuation). Soil in the bioreactor used for some experimental optional design was sterilized three times by autoclaving at 121⁰C for 15 min. All the bioreactors with its contents were incubated at room temperature (28⁰C ±2⁰C) for eight weeks under laboratory condition. The water (moisture) content of soil in each bioreactor was adjusted every week by addition of distill water in order to make up for water losses used by microbes to enhance their activities. The total bacterial counts for the treatments were carried out in representative soil composite samples using the standard serial dilution and nutrient agar-plate counting techniques (Lorch *et al.*, 1995). In order to avoid anaerobic conditions, the samples in the microcosms were tilled and aerated daily to enhance oxygenation and kept moist during the experimental period. Samples will be withdrawn at intervals of 1 week for residual total pesticide content, total heterotrophic bacteria count and other physico-chemical analysis. The experiments were carried out in triplicates.



Plate 1: wooden microcosm stacked with different treatment options as presented in Table 1

Inoculum Preparation and Enrichment of Mixed Bacterial Culture

The pure isolate (*Bacillus subtilis* and *Pseudomonas aeruginosa*) in form of slants, was obtained from veterinary research institute in Vom, Jos, Plateau State. The slant was sub-cultured using the streaking method on the nutrient agar in disposable petri dishes for 24 h in an incubator at 37°C to monitor its cell growth. A loopful of each stock culture of *Bacillus subtilis* and *Pseudomonas aeruginosa* were respectively inoculated into 3 different 250 ml Erlenmeyer flasks containing 100 ml of freshly prepared sterile nutrient broth medium (0.8%) made up of yeast extract 2.0 g/L, peptone 5.0 g/L, NaCl 5.0 g/L, and agar 15.0 g/L, incubated at 37°C for 48 hrs to monitor a turbid suspension. 1 ml of the stock solution was serially diluted into 9 ml of sterile nutrient broth (same constituents above) in a serial dilution in triplicate for each stock solution to get a viable count of 30-300 cfu/ml. 200 milliliters (2.3×10^5 CFU/g *Bacillus subtilis*, 2.5×10^5 CFU/g *Pseudomonas aeruginosa* and 2.6×10^5 CFU/g Combine *Bacillus subtilis* & *Pseudomonas aeruginosa*) of the inoculum was inoculated on the surface of the microcosm and was also used for the bioaugmentation experimental design option to study the biodegradation of glyphosate in soil. All the microcosms were mixed three times every week for aeration for the eight (8) weeks. After mixing, these microcosms were kept away from sunlight at room temperature in order to prevent rate of dehydration.

Extraction and Determination of Glyphosate Residual Content

5 g each of soil contaminated with organophosphate pesticide was weighed on an electronic weighing balance and transferred into a centrifuge tube with screw caps; 5 ml of n-hexane was added. The samples were vortexed briefly for 5 min, the mixtures were further shaken using a mechanical shaker for 10 min at room temperature. After settling, the supernatants were transferred into a clean pre-weighed 50 ml white reagent bottle. This procedure was repeated three times to bring the total solvent volume to 20 ml. The supernatant was allowed to settle and then decanted into the 50 ml white reagent bottle. The extract in the bottle was then allowed to settle for a period of 24 hours after which part of the extract was taken for the UV-VIS spectrophotometric analysis to measure the absorbance and concentration for the pesticide residual content at an optimum wavelength of 390 nm.

Statistical Analysis

Mean concentrations of pollutant with different bioremediation strategies were statistically treated with Microsoft Excel 2010. One-way between subjects ANOVA (Analysis of variance) was used to determine if there was a statistically significant difference in mean concentration of the treatment options within and between groups in the degradation of the organophosphate pesticide in the contaminated soil sample.

III. RESULTS AND DISCUSSION.

Microbial Analysis

Addition of a consortium of *Pseudomonas aeruginosa* and *Bacillus subtilis* on pesticide-polluted soil offers an interesting possibility of enhancing decontamination of the soil after a few weeks. Counts at the end of the investigation revealed that sample A4 in the organophosphate pesticide-contaminated soil treated with

combined biostimulation and bioaugmentation strategies supported more bacteria growth. Figure 1 shows the variation of Total Heterotrophic Bacteria Counts (THBC) against bioremediation time, the counts are in order of A4> A3>A2> A1 with maximum count (A4) obtained at week 6. This indicated that in the soil, the addition of preselected bacteria (bioaugmentation) and nutrients (biostimulation) resulted in the highest number of bacteria at the beginning of remediation period as compared to natural attenuation (or bioattenuation). The lower bacteria population in natural attenuation treatment may be due to the presence of low nutrients (limiting nutrient) and the disturbance of nutrient ratio especially of C:N:P that is needed in the correct ratio for bacteria growth.

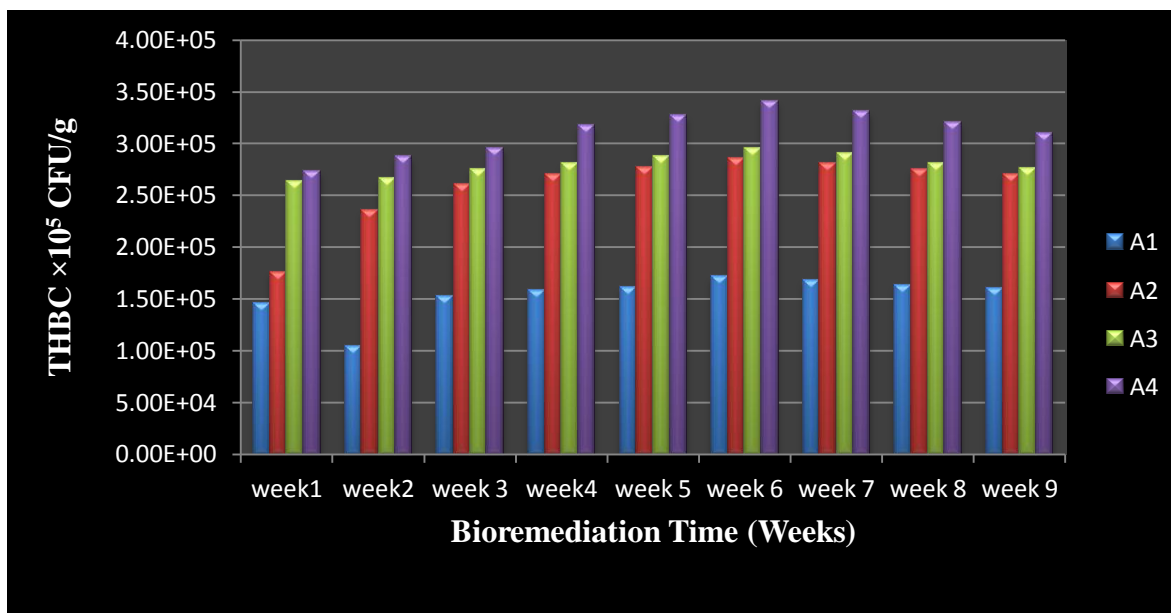


Figure 1: Variation of THBC with Bioremediation Time

The variation in concentrations of pesticide residual content (mm/kg and %) against bioremediation time is shown in Figure 2, biodegradation of the organophosphate pesticide started very fast during the first week of remediation in all the treatments and slowly continued up to the sixth week (day 42). The concentration of glyphosate was reduced from the initial concentration of 50000mg/kg to 29000 mg/kg, 20300 mg/kg, 16400 mg/kg, and 6500 mg/kg in six (6) weeks of remediation and corresponding to 42%, 59.4%, 67.2%, and 87% glyphosate reduction was achieved under A1, A2, A3 and A4 respectively. This observation revealed that during the glyphosate biodegradation in soil, addition of organic manure (Fortified Cow Dung) and bacterial inoculums individually resulted in a more effective bioremediation response than the natural attenuation. Similar observations have been reported for, naphthalene (Agarry and Oghenejoboh, 2015) and spent engine oil (Abdulsalam and Omale; 2009 and Abdulsalam et al., 2011). Nevertheless, bioaugmentation strategy elicited a higher biodegradation of glyphosate than the biostimulation treatment. This is in agreement with the observation of (Bento et al., (2003) and Agarry and Oghenejoboh, 2015) who reported that among the individual methods of natural attenuation, biostimulation, and bioaugmentation that were used for the remediation of a soil contaminated by diesel oil, bioaugmentation method elicited higher diesel oil degradation than others. In contrast, other workers have shown that biostimulation strategy enhanced the bioremediation of kerosene contaminated soil (Shabir et al., 2008; Agarry and Oghenejoboh, 2015), crude oil contaminated soil (Xu and Lu, 2010), spent engine oil contaminated soil (Abdulsalam et al.,2011) and lubricating oil contaminated soil (Agarry et al., 2013) than bioaugmentation. Generally, in this work, the combination of biostimulation (Organic Manure) and bioaugmentation treatment strategy (which has not been reported for glyphosate removal) showed relatively greater glyphosate reduction than other treatments during the whole period of remediation.

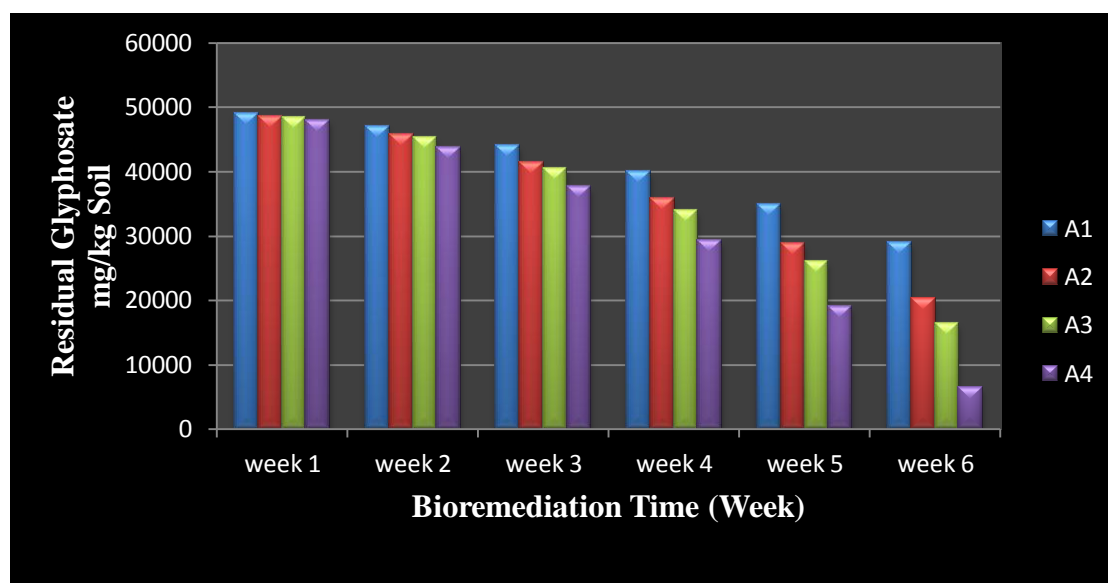


Figure 2: Variation of Residual Glyphosate content with Bioremediation Time.

IV. Conclusion

The work on enhanced aerobic biodegradation of organophosphate pesticide in soil was carried out successfully within the limits of experimental errors. The following conclusions can be drawn based on the findings from the work. The hybrid of bioaugmentation and biostimulation exhibited the highest degree of degradation at their various mean rate of the extent of percentages in pesticide concentrations removal, followed by Bioaugmentation treatment Options, then the soil treatment with Organic manure(cattle Dung) Biostimulation options and natural attenuation been the least removal rate of Organophosphate pesticide percentage concentration removal rate (combined Biostimulation and Bioaugmentation> Bioaugmentation> Biostimulation> Natural Attenuation). This same trend was also observed with the THBC analysis with the hybrid of bioaugmentation and biostimulation (A7) having the highest number of the bacterial count in the highest Exponential phase of the analysis at week 6. The low THBC counts in other treatment options could be attributed to availability of nutrients between the exogenous and native microbes. Therefore A7 was the best in cleaning OPP contaminated soil artificially.

References

- [1]. Abdulsalam, S., Bugaje, I. M., Adefila, S. S., Ibrahim, S. (2011) Comparison of biostimulation and bioaugmentation for remediation of soil contaminated with spent motor oil. *International Journal of Environmental Science and Technology* 8(1), 187-194, 2011
- [2]. Abdulsalam, S., Omale, A.B (2009).. "Comparison of biostimulation and bioaugmentation techniques for the remediation of used motor oil contaminated soil". *Brazilian Archives of Biology and Technology* 52, 747-754.
- [3]. Agarry, S. E., Oghenejoboh, K. M. (2015). "Enhanced Aerobic Biodegradation of Naphthalene in Soil: Kinetic Modeling and Half-Life Study". *International Journal of Environmental Bioremediation & Biodegradation*, Vol. 3, No. 2, 48-53 Available online at <http://pubs.sciepub.com/ijebb/3/2/2> © Science and Education Publishing DOI:10.12691/ijebb-3-2-2
- [4]. Agarry, SE, Aremu, MO., Aworanti, OA. (2013) Kinetic modelling and half-life study on bioremediation of soil co-contaminated with lubricating motor oil and lead using different bioremediation strategies. *Soil and Sediment Contamination- An International Journal* 22 (7), 800-816, 2013
- [5]. Asogwa, E.U and Dongo, L.N (2009). Problem associated with pesticide usage and application in Nigerian Cocoa production: a review. *Afri. j. agri. Res.* 4(8): 675-683
- [6]. Bento, F. M., Camargo, F.A., Okeke, B., Frankenberger, Jr. T.W. (2003) Bioremediation of soil contaminated by diesel oil. *Brazilian Journal of Microbiology* 34(Suppl. 1), 65-68, 2003.
- [7]. Changpeng Zhang , Xiuqing Hu , Jinyan Luo , Zhiyi Wu , Li Wang , Bin Li , Yanli Wang and Guochang Sun. (2015). Degradation Dynamics of Glyphosate in Different Types of Citrus Orchard Soils in China. *Molecules* 2015, 20, 1161-1175; doi: 10.3390/molecules20011161
- [8]. He, H.M.; Zhang, C.R.; Zhu, Y.H.; Zhang, C.P.; Ping, L.F.; Zhao, H.; Wu, M.; Tang, T.; Cai X.M.; Li, Z. (2014). Residue and degradation of cyantraniliprole and its main metabolite in pepper and soil. *Chin. J. Anal. Chem.* vol. 42, 1177–1182
- [9]. Kang, D.G., Choi., S.S. and Cha, H.J. (2006). Enhanced biodegradation of toxic organophosphate compounds using recombinant *Escherichia coli* with sec pathway driven periplasmic secretion of organophosphorous hydrolase. *Biotechnol. Prog* 22: 406-410
- [10]. Li, Y.F.; Zhang, C.; Yin, Y.H.; Cui, F.; Cai, J.Y.; Chen, Z.H.; Jin, Y.H.; Robson, M.G.; Li, M.; Ren, Y.T.; (2014). Neurological effects of pesticide use among farmers in China. *Int. J. Environ. Res. Public Health* vol. 11, 3995–4006.
- [11]. Mink, P.J.; Mandel, J.S.; Scurman, B.K.; Lundin, J.I. (2012). Epidemiologic studies of glyphosate and cancer: A review. *Regul. Toxicol. Pharmacol.*, vol. 63, 440–452.
- [12]. Nwogu, T.P, Azubuikwe, C. C. and Ogugbue, C. J. (2015). Enhanced Bioremediation of Soil Artificially Contaminated with Petroleum Hydrocarbon after Amendment with *Capra aegagrus hircus* (Goat) Manure. *Biotechnology Research International* vol, 2015, Article ID 657349, 7 pages

- [13]. Schönbrunn, E.; Eschenburg, S.; Shuttleworth, W.A.; Schloss, J.V.; Amrhein, N.; Evans, J.N.; Kabsch, W. (2001). Interaction of the herbicide glyphosate with its target enzyme 5-enolpyruvylshikimate 3-phosphate synthase in atomic detail. *Proc. Natl. Acad. Sci. USA*, vol. 98, 1376–1380.
- [14]. Shabir, G., Afzal, M., Anwar, F., Tahseen, R., Khalid, Z.M. “Biodegradation of kerosene in soil by a mixed bacterial culture under different nutrient conditions”. *International Journal of Biodeterioration and Biodegradation* 61, 161-166, 2008
- [15]. Steinrücken, H.C.; Amrhein, N. (1980). The herbicide glyphosate is a potent inhibitor of 5-enolpyruvyl-shikimic acid-3-phosphate synthase. *Biochem. Biophys. Res. Commun.*, vol. 94, 1207–1212
- [16]. Stephen, O.D.; Stephen, B.P. (2008) Glyphosate: A once-in-a-century herbicide: Mini-review. *Pest Manag. Sci.*, vol. 64, 319–325
- [17]. Xu, Y., Lu, M. (2010) “Bioremediation of crude oil-contaminated soil: comparison of different biostimulation and bioaugmentation treatments”. *Journal of Hazardous Materials* 183 (1-3), 395-401.
- [18]. Yamada, T.; Kremer, R.J.; de Camargo e Castro, P.R.; Wood, B.W. (2009). Glyphosate interactions with physiology, nutrition, and diseases of plants: Threat to agricultural sustainability? *Eur. J. Agron.* vol., 31, 111–113
- [19]. Zhang, C.P.; Zhao, H.; Cai, X.M.; He, H.M.; Zhu, Y.H.; Li, Z. (2012) Residue analysis and degradation dynamics of tebuconazole in rice. *Agrochemicals* vol, 51, 675–679