



Chemical constituents of the leaves of *Aralia hiepihana*

Nguyen Thi Thu Hien^{1,2}, Nguyen Huu Huong Duyen¹, Nguyen Thi Dieu Thuan¹, Tran Thi Ngoc Hanh¹, Pham Van Huyen¹, Nguyen Huu Toan Phan^{1,2*}

¹Tay Nguyen Institute for Scientific Research, VAST, 116 Xo Viet Nghe Tinh, Dalat, Vietnam

²Graduate University of Science and Technology, VAST, 18 Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam

Corresponding Author: Nguyen Huu Toan Phan

ABSTRACT: Phytochemical study of the leaves of *Aralia hiepihana*, which is an endemic species of Lam Dong province, Vietnam, led to the isolation of six compounds, named quercetin-3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside (**1**), rutin (**2**), ursolic acid (**3**), methyl 3,4 dihydroxybenzoate (**4**), methyl caffeate (**5**), and araliasaponin IV (**6**). Their structures were elucidated by 1D, 2D NMR and MS spectroscopic analyses in comparison with the data reported in the literature. These metabolites isolated for the first time from this species.

KEYWORDS: *Aralia hiepihana*, ursolic acid, methyl caffeate, rutin, quercetin-3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside, araliasaponin IV.

Received 20 August, 2020; Accepted 06 September, 2020 © The author(s) 2020.

Published with open access at www.questjournals.org

I. INTRODUCTION

Aralia genus belongs to the Araliaceae family, consisting of 79 accepted species of deciduous or evergreen trees, shrubs, and rhizomatous herbaceous perennials, distributed in the Asia and America. Some *Aralia* species have been used in the treatment of respiratory inflammation, diabetes, cancer, liver protective and parasitic infections [1-7].

Aralia hiepihana J. Wen & Lowry, an endemic species of southern Vietnam was described and illustrated in 2002 [8]. In the previous report, five flavonoid compounds were isolated from the leaves of this species [9]. Continuing our research, six compounds **1-6** have been isolated for the first time from the leaves of *A. hiepihana*.

II. EXPERIMENTS

2.1. General experimental procedures

Column chromatography (CC) was performed on silica gel (Kieselgel 60, 70–230 mesh and 230–400 mesh, Merck, Darmstadt, Germany) and reversed-phase silica gel (ODS-A, 12 nm S-150 mm, YMC Co., Ltd., Japan) resins. TLC used pre-coated silica gel 60 F₂₅₄ (1.05554.0001, Merck) and RP-18 F_{254S} plates (1.15685.0001, Merck), and compounds were visualized by spraying with aqueous 10% H₂SO₄ and heating for 3–5 min. The ¹H NMR (500 MHz) and ¹³C NMR (125 MHz) spectra were recorded on an AVANCE III HD 500 (Bruker, Germany) FT-NMR spectrometer with tetramethylsilane (TMS) was used as an internal standard. ESI mass spectra were collected on Agilent 1100 LC/MS systems.

2.2. Plant material

The leaves of *A. hiepihana* were collected at Dalat, Lamdong, Vietnam in 2017 and identified by Dr. Nong Van Duy, Tay Nguyen Institute for Scientific Research, VAST. A voucher specimen (No.TN3/129) is deposited at the Herbarium of Tay Nguyen Institute for Scientific Research, VAST, Vietnam.

2.3. Extraction and isolation

The powdered leaves of *A. hiepihana* (4.5 kg) were extracted three times with methanol at room temperature and evaporated under low pressure to obtain 619 g residue. This residue was suspended in distilled water (2 L) and partitioned in turn with *n*-hexane, CHCl₃, and EtOAc to give corresponding extracts *n*-hexane (H, 187 g), CHCl₃ (C, 17 g), EtOAc (E, 52 g), and water layer (W, 2L).

The extract E (52 g) was separated on a silica gel chromatography column (CC), eluted with CHCl₃-MeOH (100:0 - 0:100, v/v) to yield five fractions, E1-E5. Fraction E4 (23 g) was fractionated on a silica gel CC eluted with CH₂Cl₂-acetone-MeOH (10:1:1, 5:1:1, 3:1:1, v/v/v) to give ten subfractions, E4.1-E4.10. Subfraction E4.5 (5.4 g) was purified on a silica gel CC using CH₂Cl₂-acetone (6:1, v/v) to afford compound **3** (1.0 g). Fraction E5 (30 g) was separated on a silica gel CC with CHCl₃-MeOH (100:0 - 0:100, v/v) to get fourteen subfractions, E5.1-E5.14. Subfraction E5.6 (12 g) was separated on a silica gel CC with *n*-hexan-EtOAc (1:1, v/v) to furnish five subfractions, E5.6.1- E5.6.5. Subfraction E5.6.2 (504 mg) further purified on silica gel CC with CH₂Cl₂-acetone (15:1, v/v) followed by ODS-A CC with MeOH-H₂O (1:2.5, v/v) to give compound **4** (16 mg) and compound **5** (28 mg).

The W layer was passed through Diaion HP-20 CC using step-wise eluent of MeOH-H₂O (0:100, 25:75, 50:50, 75:25, and 100:0, v/v) to obtain four fractions W1-W4, after removal of the fraction eluted with 100% water. Fraction W2 (61 g) was separated on silica gel CC with a gradient mixture of CHCl₃-MeOH (100:0 - 0:100, v/v) to obtain ten fractions, W2.1-W2.10.

Fraction W2.6 (9.5 g) was further separated into ten subfractions, W2.6.1-W2.6.10, using silica gel CC with eluent EtOAc-MeOH-H₂O (30:10:2, v/v/v). Subfraction W2.6.10 (1.7 g) was passed through to Sephadex LH-20 CC with MeOH-H₂O (2:1, v/v), followed by silica gel CC with CHCl₃-MeOH (3:1, v/v), afforded compound **6** (15 mg).

Fraction W2.10 (6.8 g) was further applied to Sephadex LH-20 CC using a gradient of MeOH-H₂O (1:3 - 1:0, v/v) to obtain five subfractions. Subfraction W2.10.2 (5.0 g) was purified by silica gel CC with CHCl₃-MeOH (2:1 v/v), followed by ODS-A CC with MeOH-H₂O (1:1, v/v), yielding compounds **1** (10 mg) and **2** (5 mg).

Quercetin-3-O-β-D-glucopyranoside-7-O-α-L-rhamnopyranoside (1): Yellow powder, molecular formula C₂₇H₃₀O₁₆, ESI-MS: *m/z* 611 [M+H]⁺ and *m/z* 609 [M-H]⁻. ¹H NMR (500 MHz, CD₃OD) and ¹³C NMR (125 MHz, CD₃OD) see table 1.

Rutin (2): Light yellow powder, molecular formula C₂₇H₃₀O₁₆, ESI-MS: *m/z* 611 [M+H]⁺ and *m/z* 609 [M-H]⁻. ¹H NMR (500 MHz, CD₃OD) and ¹³C NMR (125 MHz, CD₃OD) see table 1.

Ursolic acid (3): White amorphous powder, molecular formula C₃₀H₄₈O₃, ESI-MS: *m/z* 457.2 [M+H]⁺ and *m/z* 455.3 [M-H]⁻. ¹H-NMR (500 MHz, CD₃OD) δ_H: 5.25 (1H, t, *J* = 3.5 Hz, H-12), 3.18 (1H, dd, *J* = 4.5, 11.0 Hz, H-3), 2.22 (1H, d, *J* = 11.0 Hz, H-18), 0.98 (3H, s, H-27), 0.99 (3H, s, H-23), 1.13 (3H, s, H-30), 0.98 (3H, s, H-25), 0.90 (3H, s, H-29), 0.87 (3H, s, H-26); 0.80 (3H, s, H-24). ¹³C-NMR (125 MHz, CD₃OD) δ_C: 40.01 (C-1), 29.21 (C-2), 79.73 (C-3), 39.83 (C-4), 56.75 (C-5), 19.48 (C-6), 34.34 (C-7), 40.79 (C-8), 49.04 (C-9), 38.00 (C-10), 24.36 (C-11), 126.91 (C-12), 139.63 (C-13), 43.25 (C-14), 27.89 (C-15), 25.32 (C-16), 48.49 (C-17), 54.37 (C-18), 40.40 (C-19), 40.42 (C-20), 31.76 (C-21), 38.10 (C-22), 28.78 (C-23), 16.38 (C-24), 16.02 (C-25), 17.81 (C-26), 21.56 (C-27), 181.66 (C-28), 17.64 (C-29), 24.10 (C-30).

Methyl 3,4-dihydroxybenzoate (4): Brown amorphous powder, molecular formula C₈H₈O₄, ESI-MS: *m/z* 169.1 [M+H]⁺. ¹H NMR (500 MHz, CD₃OD) δ_H: 7.44 (1H, d, *J* = 2.0 Hz, H-2), 7.43 (1H, dd, *J* = 2.0, 8.0 Hz, H-6), 6.81 (1H, d, *J* = 8.0 Hz, H-5), 3.84 (3H, s, -OCH₃). ¹³C NMR (125 MHz, CD₃OD) δ_C: 122.60 (C-1), 117.42 (C-2), 146.15 (C-3), 151.67 (C-4), 115.85 (C-5), 123.63 (C-6), 168.86 (C-7), 52.22 (OCH₃).

Methyl caffeate (5): White amorphous powder, molecular formula C₁₀H₁₀O₄, ESI-MS: *m/z* 195.1 [M+H]⁺. ¹H NMR (500 MHz, CD₃OD) δ_H: 7.55 (1H, d, *J* = 16.0 Hz, H-7), 7.05 (1H, d, *J* = 2.0 Hz, H-2), 6.95 (1H, d, *J* = 2.0, 8.0 Hz, H-6), 6.79 (1H, d, *J* = 8.0 Hz, H-5), 6.26 (1H, d, *J* = 16.0 Hz, H-8), 3.77 (3H, s, -OCH₃). ¹³C NMR (125 MHz, CD₃OD) δ_C: 127.70 (C-1), 115.15 (C-2), 146.77 (C-3), 149.52 (C-4), 116.49 (C-5), 122.89 (C-6), 146.90 (C-7), 114.85 (C-8), 168.86 (C-9), 51.96 (OCH₃).

Araliasaponin IV (6): White amorphous powder, molecular formula C₅₃H₈₆O₂₂, ESI-MS [M+Na]⁺ at *m/z* 1098. ¹H NMR (500 MHz, pyridine-*d*₅): δ_H: 3.20 (1H, dd, *J* = 4.0, 11.0 Hz, H-3), 5.38 (1H, m, H-12), 3.13 (1H, dd, *J* = 3.5, 13.5 Hz, H-18), 1.20 (1H, s, H-23), 1.05 (1H, s, H-24), 0.80 (1H, s, H-25), 1.05 (1H, s, H-26), 1.20 (1H, s, H-27), 0.83 (1H, s, H-29), 0.83 (1H, s, H-30), 4.76 (1H, d, *J* = 7.5 Hz, H-1'), 5.37 (1H, d, *J* = 8.0 Hz, H-1''), 5.22 (1H, d, *J* = 8.0 Hz, H-1'''), 6.22 (1H, d, *J* = 8.0 Hz, H-1'''). ¹³C NMR (125 MHz, CD₃OD) δ_C: 38.63 (C-1), 28.07 (C-2), 89.32 (C-3), 39.50 (C-4), 55.78 (C-5), 18.38 (C-6), 32.35 (C-7), 39.74 (C-8), 47.87 (C-9), 36.69 (C-10), 23.24 (C-11), 122.73 (C-12), 144.00 (C-13), 42.00 (C-14), 26.48 (C-15), 23.48 (C-16), 46.89 (C-17), 41.59 (C-18), 46.09 (C-19), 30.59 (C-20), 33.83 (C-21), 32.96 (C-22), 27.66 (C-23), 16.34 (C-24), 15.38 (C-25), 17.29 (C-26), 25.94 (C-27), 176.49 (C-28), 32.96 (C-29), 23.62 (C-30), 105.20 (C-1'), 77.11 (C-2'), 84.77 (C-3'), 69.25 (C-4'), 75.75 (C-5'), 61.89 (C-6'), 104.63 (C-1''), 77.94 (C-2''), 78.43 (C-3''), 71.07 (C-4''), 66.76 (C-5''), 104.97 (C-1'''), 75.81 (C-2'''), 79.02 (C-3'''), 70.83 (C-4'''), 78.59 (C-5'''), 62.05 (C-6'''), 95.55 (C-1''''), 73.77 (C-2''''), 75.03 (C-3''''), 71.25 (C-4''''), 78.06 (C-5''''), 62.17 (C-6'''').

III. RESULTS AND DISCUSSION

Compound **1** was isolated as yellow powder. The molecular formula was established as $C_{27}H_{30}O_{16}$ by ESI-MS data ($[M+H]^+$ m/z 611 and $[M-H]^-$ m/z 609). The 1H NMR spectrum showed the signals of two AX-type aromatic protons [δ_H 6.79 (d, $J = 2.0$ Hz, H-8) and 6.50 (d, $J = 2.0$ Hz, H-6)] and three protons of an ABX system [δ_H 7.86 (d, $J = 2.5$ Hz, H-2'), 7.65 (dd, $J = 2.5, 8.5$ Hz, H-6'), and 6.90 (d, $J = 8.5$ Hz, H-5')]. Moreover, the signals of two anomeric protons at δ_H 5.59 (d, $J = 1.5$ Hz, H-1''') and 5.25 (d, $J = 8.0$ Hz, H-1'') corresponded to two anomeric carbons at δ_C 99.93 and 105.04 were assigned to α -L-rhamnose (Rha) and β -D-glucose (Glc) units, respectively. The ^{13}C NMR and DEPT spectrum confirmed the presence of twenty seven carbons including one carbonyl carbon at δ_C 179.75 (C-4), nine quaternary carbons, seven methine carbons, eight oxymethine carbons, one oxymethylene carbon, and one methyl carbon. Therefore, the aglycon of **1** was identified as quercetin. The HMBC spectrum showed correlations between anomeric proton at δ_H 5.25 (H-1'') of Glc and carbon at δ_C 135.98 (C-3) of aglycon, between anomeric proton at δ_H 5.59 (H-1''') of Rha and carbon at δ_C 163.68 (C-7), which suggest the glycosylation at C-3 and C-7 of quercetin skeleton. Based on data of ESI-MS, 1D, 2D NMR and compared with previous published data [10], the structure of **1** was determined as quercetin 3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside.

Compound **2** was also obtained as a yellow powder and its molecular formula, $C_{27}H_{30}O_{16}$, was determined by ESI-MS with a quasi-molecular ion peak at m/z 611 $[M+H]^+$. Its NMR data were similar to those of **1** (Table 1) including signals of quercetin, Rha, and Glc units. The 1H NMR spectra of **2** also showed two anomeric proton signals at δ_H 5.13 (d, $J = 7.5$ Hz, H-1'') and 4.54 (d, $J = 1.0$ Hz, H-1''') were assigned to α -L-rhamnose (Rha) and β -D-glucose (Glc) units, respectively. The position of attachment of the sugar moiety was obtained from the HMBC spectrum, in which correlations were seen between the anomeric proton of glucose H-1'' and carbon of the aglycon (C-3). In addition, a cross-peak between the signal at δ_H 4.54 (H-1''') and δ_C 68.56 (C-6'' of the glucose) confirmed that the glycosylation of the glucose unit by the rhamnose took place on the 6''-hydroxyl. Therefore, compound **2** was identified as rutin [11].

Table 1: The NMR data of **1** and **2**

Position	1		2	
	δ_C	δ_H (J in Hz)	δ_C	δ_H (J in Hz)
2	159.39		158.52	
3	135.98		135.63	
4	179.75		179.43	
5	162.83		162.99	
6	100.62	6.50 d. 2.0	99.96	6.23 d. 2.0
7	163.68		166.04	
8	95.56	6.79 d. 2.0	94.87	6.42 d. 2.0
9	158.1		159.34	
10	107.37		105.64	
1'	122.75		123.14	
2'	117.77	7.86 d. 2.5	117.70	7.69 d. 2.0
3'	145.89		145.85	
4'	150.17		149.81	
5'	116.17	6.90 d. 8.5	116.07	6.90 d. 8.5
6'	123.17	7.65 dd. 2.5. 8.5	123.56	7.65 dd. 2.0. 8.5
1''	105.04	5.25 d. 8.0	104.72	5.13 d. 7.5
2''	73.62	3.49 m	75.73	3.49 m
3''	75.06	3.57 m	78.20	3.44 m
4''	71.29	3.62 m	71.41	3.29 m
5''	77.26	3.49 m	77.23	3.35 m
6''	62.02	H _a : 3.65 dd. 6.0; 11.0 H _b : 3.58 m	68.56	H _a : 3.82 dd. 1.0; 11.0 H _b : 3.41 m
1'''	99.93	5.59 d. 1.5	102.42	4.54 (d. 1.0)
2'''	70.09	3.87 m	72.11	3.66 m
3'''	71.71	4.04 m	72.25	3.56 dd
4'''	73.95	3.49 m	73.95	3.30 m*
5'''	73.17	3.83 m	69.71	3.47 m
6'''	18.05	1.27 d. 6.0	17.88	1.14 d. 6.0

Compound **3** was obtained as white amorphous powder. The molecular formula was established as $C_{30}H_{46}O_3$ by ESI-MS data ($[M+H]^+$ m/z 457.2 and $[M-H]^-$ m/z 455.3). The ^{13}C NMR and DEPT spectrum indicated that **3** has total 30 carbons including seven methyls, nine methylenes, six sp^3 methines, one sp^2 methine, five quaternary sp^3 carbons, and two quaternary sp^2 carbons. The three sp^2 carbons (δ_C 126.91, 139.63, and 181.66) indicated a double bond and carbonyl group of the ursane-type triterpenoid acid. The 1H NMR spectrum also showed signals of seven methyl groups [δ_H 1.17 (s, H-23), 0.94 (s, H-24), 0.80 (s, H-25), 0.96 (s, H-26), 1.15 (s, H-27), 0.95 (s, H-29), 0.89 (s, H-30)], one trisubstituted olefin [δ_H 5.42 (dd, $J = 4.0, 8.5$ Hz, H-12)], and one oxymethine [δ_H 3.18 (dd, $J = 4.5, 11.0$ Hz, H-3)]. Furthermore, HMBC correlations observed from H-2 to C-1, C-3, and C-4, from H-3 to C-4, C-23, and C-24, from H-11 to C-12 and C-13, from H-29 to C-19, from H-30 to C-20, and from H-18 and H-16 to C-28 allowed to confirm the structure of **3**. Based on NMR spectroscopic data combined comparison with literature data [12] the structure of **3** was established as ursolic acid.

Compound **4** was obtained as brown amorphous powder. The 1H NMR spectra showed the signals of three protons in 1,3,4-trisubstituted benzene ring at δ_H 7.44 (1H, d, $J = 2.0$ Hz, H-2), 7.43 (1H, dd, $J = 2.0, 8.0$ Hz, H-6), and 6.81 (1H, d, $J = 8.0$ Hz, H-5) and the signal at δ_H 3.84 (3H, s) of a methoxy group in the structure. The ^{13}C NMR and DEPT spectrum confirm the presence of eight carbons including one carbonyl carbon at δ_C 168.86, two oxygenated aromatic carbons at δ_C 146.15 and 151.67, four other aromatic protons at δ_C 123.63, 122.6, 117.42, and 115.85, and one methoxy carbon at δ_C 52.22. Thus, compound **4** was recognized as 3,4-dihydroxybenzoic methyl ester with molecular formula $C_8H_8O_4$ based on the spectral data and the literature [13].

Compound **5** was obtained as a white amorphous powder. The structure of **5** very similar with the structure of **4** except for the presence of the olefinic signal at δ_H 7.55 (1H, d, $J = 16.0$ Hz, H-7) and 6.26 (1H, d, $J = 16.0$ Hz, H-8). The 1H NMR spectrum also revealed signals in a 1,3,4-trisubstituted benzene ring at δ_H 7.05 (1H, d, $J = 2.0$ Hz, H-2), 6.95 (1H, d, $J = 2.0, 8.0$ Hz, H-6), and 6.79 (1H, d, $J = 8.0$ Hz, H-5) and one methoxy group at δ_H 3.77 (3H, s). The ^{13}C and DEPT spectrum confirmed ten signals include five methin carbons, four quaternary carbons and one methoxy group. In the HMBC spectrum data determined the correlation between the proton of olefinic with C-2 (δ_C 127.7) of benzene ring, (H-1'/C-2, C-3, C-4, C-5) and H-2'/C-2, C-1', C-3'). From the above spectral data and compared with the literature [14], compound **5** was identified as methyl caffeate with its molecular formula being $C_{10}H_{10}O_4$.

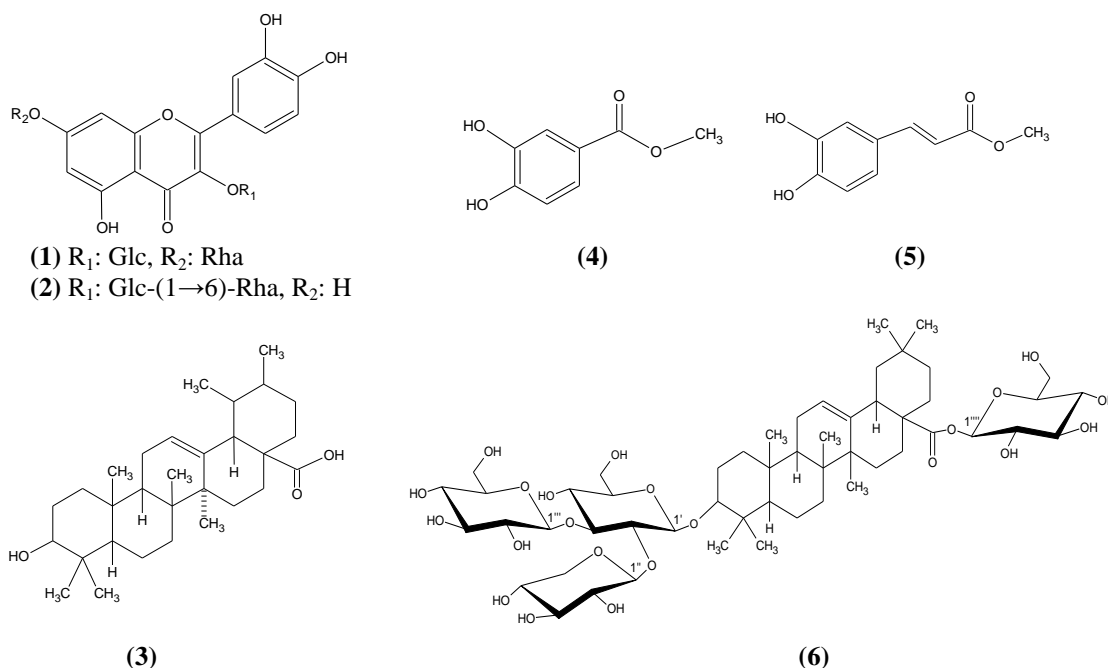


Figure 1: The structure of six compounds (1-6)

Compound **6** was isolated as a white amorphous powder. The 1H NMR spectrum showed the presence of seven singlet methyl protons δ_H 1.20, 1.05, 0.83 (each 6H, s), 0.80 (3H, s), a trisubstituted olefinic proton (δ_H 5.38 m, H-12) which was characteristic of olean-type triterpene skeleton and four anomeric proton signals [(δ_H 4.76 d, $J = 7.5$ Hz, H-1'), (δ_H 5.37, d, $J = 8.0$ Hz, H-1''), (δ_H 5.22, d, $J = 8.0$ Hz, H-1'''), and (δ_H 6.22, d, $J = 8.0$ Hz)] of the sugar moieties. The ^{13}C NMR spectrum showed the presence of an ester carbonyl carbon (δ_C 176.46 (C-28) and five anomeric carbon signals [(δ_C 105.20 (Glu, C-1'), 104.63 (Xyl, C-1''), 104.97 (Glu, C-1'''), and 95.55 (Glu, C-1''')]. In the HMBC, the correlations were observed between the proton signal at δ_H 4.76 (Glu, H-1') to the carbon signal at δ_C 89.32 (C-3) of the aglycon and between the anomeric proton signal at δ_H 5.37 (Xyl,

H-1^{''}) and the carbon signal at δ_C 77.11 (Glu-C-2'), the anomeric proton signal at δ_H 5.22 (Glu, H-1^{'''}) and the carbon signal at δ_C 84.77 (Glu-C-3'). In addition, an anomeric proton at δ_H 6.22 (Glu-1^{''''}) showed the correlation with the carbon signal at δ_C 176.46 (C-28) which suggested glycosylation at C-3 of aglycon with a ([xyl-(1 \rightarrow 2)]-[glc-(1 \rightarrow 3)]-glc) moiety and at C-28 with a glucose unit. Accordingly, the structure of **6** was elucidated as araliasaponin IV, which was isolated from *A. decaisneana* [15] and *Catunaregam spinosa* [16].

IV. CONCLUSION

From the leaves of *A. hiepinana* using combined chromatographic methods obtained six compounds, including quercetin-3-O- β -D-glucopyranoside-7-O- α -L-rhamnopyranoside, rutin, ursolic acid, methyl 3,4-dihydroxybenzoate, methyl caffeate, and araliasaponin IV. These structures were elucidated by 1D, 2D NMR, and mass spectrum. All of them were isolated for the first time from this plant.

Acknowledgement: This work was financially supported by Vietnam national project of the Tay Nguyen 3 Program in the period 2016-2020 (code: TN18/C09).

REFERENCES

- [1]. Yoshikawa, M., Murakami, T., Harada, E., Murakami, N., Yamahara, J., Matsuda, H., Bioactive Saponins and Glycosides. VII. On the hypoglycemic principles from the root cortex of *Aralia elata* SEEM.: Structure Related Hypoglycemic Activity of Oleanolic Acid Oligoglycoside. Chem. Pharm. Bull., 1996. **44**: p. 1923-1927.
- [2]. Wang, Z., Hu, J., Su, Z., Li, C., Li, R., Tang, H., Yi, Y., Liver-protective activity of *Aralia taibaiensis*. Zhongguo Zhong Yao Za Zhi, 1997. **22**(5): p. 307-308.
- [3]. Tan, B. K. and Vanitha, J., Immunomodulatory and antimicrobial effects of some traditional chinese medicinal herbs: a review. Curr Med Chem, 2004. **11**(11): p. 1423-1430.
- [4]. Bhat, Z. A., Ansari, S. H., Mukhtar, H. M., Naved, T., Siddiqui, J. I., Khan, N. A., Effect of *Aralia cachemirica* Decne root extracts on blood glucose level in normal and glucose loaded rats. Pharmazie, 2005. **60**(9): p. 712-713.
- [5]. Lee, I. S., Jin, W.Y., Zhang, X., Hung, T. M., Song, K. S., Y Seong, Y. H., Bae, K., Cytotoxic and COX-2 inhibitory constituents from the aerial parts of *Aralia cordata*. Arch Pharm Res, 2006. **29**(7): p. 548-555.
- [6]. Qi, L-W., Liu, E-H., Chu, C., Peng, Y-B., Cai, H-X., Liet, P., Anti-diabetic agents from natural products an update from 2004 to 2009. Curr Top Med Chem, 2010. **10** (4): p. 434-457.
- [7]. Dou, F., Xi, M., Wang, J., Tian, X., Hong, L., Tang, H., Wen, A., Glucosidase and amylase inhibitory activities of saponins from traditional Chinese medicines in the treatment of diabetes mellitus. Pharmazie, 2013. **68**: p. 300-304.
- [8]. Wen, J. and Lowry, P., *Aralia hiepinana* J. Wen & Lowry, a new species of Araliaceae from Vietnam. Adansonia III, 2002. **24**(2): p. 213-216.
- [9]. Thuan, N. T. D., Hien, N. T. T., Hao, T. M., Huyen, P. V., Phan, N. H. T., Flavonoids from the leaves of *Aralia hiepinana*. Vietnam J. Sci. Tech., 2018. **56**(4A): p. 259-265.
- [10]. Kim, H. J., Kim, B.-G., and Ahn, J.-H., Regioselective synthesis of flavonoid bisglycosides using *Escherichia coli* harboring two glycosyltransferases. Appl Micro. Biotech., 2013. **97**: p. 5275-5282.
- [11]. Zhou, X., Peng, J., Fan, G., Wu, Y., Isolation and purification of flavonoid glycosides from *Trollius ledebouri* using high-speed counter-current chromatography by stepwise increasing the flow-rate of the mobile phase. Journal of Chrom. A, 2005. **1092**: p. 216-221.
- [12]. Deng, J.-Z., Starck, S. R., and Hecht, S. M., DNA Polymerase a Inhibitors from *Baeckea gunniana*. J. Nat. Prod., 1999. **62**: p. 1624-1626.
- [13]. Sharma, N., Sharma A, Bhatia G, Landi M, Brestic M, Singh B, Singh J, Kaur S, Bhardwaj R., Isolation of Phytochemicals from *Bauhinia variegata* L. Bark and Their In Vitro Antioxidant and Cytotoxic Potential. Antioxidants, 2019. **8**: p. 492.
- [14]. Xiang, M., Su, H., Hu, J., and Yan, Y., Isolation, identification and determination of methyl caffeate, ethyl caffeate and other phenolic compounds from *Polygonum amplexicaule* var. sinense. J. Med. Pl. Res., 2011. **5**(9): p. 1685-1691.
- [15]. Miyase, T., Shiokawa, K. I., Zhang, D. M., and Ueno, A. Araliasaponins I-XI, triterpene saponins from the roots of *Aralia decaisneana*. Phytochem., 1996. **41**(5): p. 1411-1418.
- [16]. Gao, G., Lu, Z., Tao, S., Zhang, S., and Wanga, F., Triterpenoid saponins with antifeedant activities from stem bark of *Catunaregam spinosa* (Rubiaceae) against *Plutella xylostella* (Plutellidae). Car. Res., 2011. **346**: p. 2200-2205.

Nguyen Thi Thu Hien, et. al. "Chemical constituents of the leaves of *Aralia hiepinana*" Quest Journals Journal of Research in Pharmaceutical Science, vol. 06, no. 03, 2020, pp. 01-05