



Research Paper

Microbial Production from Fruit and Floral Waste: A Review

¹GandhiDharakumari,²NehaTarpura

¹M.scmicrobiologyStudent,²TeachingAssistant

^{1,2}DepartmentofMicrobiology,BhagwanMahavirCollegeofBasicandAppliedScience,BHAGWANMAHAVIR
UNIVERSITY, SuratIndia-395007

ABSTRACT:

The citrus peels contain cyclic monoterpene limonene. Limonene is the major component in the oil of citruspeel. In orange oil greater than 90% D-limonene are present. D-limonene extract by the distillation methodfromorangepeel(AmanPandayetal.,2015).AccordingtoRivasetal.,theorangepeelisinfactconstitutedbysolublesugars-16.9%,starch-3.75%,fiber(cellulose-9.21%,hemicelluloses-10.5%,lignin-0.84%andpectin-42.5%),ashes-3.50%,fats-1.95 % andproteins-6.50 % (Rivas et al., 2008).Citric acid is an intermediate of the tricarboxylic acid cycle (TCA). The first patent for citric acid production by *Aspergillus niger* utilising sugarsolutions was reported in 1913 (Tornado et al., 2011).Orange peels contains soluble sugars and pectin as themain components. According to Rivas et al. (2006).The large amount of this waste is still dumped every year,which causes both economic and environmental problems such as high transportation cost, lack of dumpingsite, andaccumulationofhighorganiccontentmaterial. Therefore,moreeffectiveandsustainablealternativesforusingorangepeelwastes(Wikandarietal.,2015;GrohmannandBaldwin,1992; Marinetal.,2007).

Rose is well known plant for aroma production and has number of industrial uses.2-phenylalcohols are one ofthemostimportantaromaticalcoholstogiveroselikeAromainfoodproducts.Thehigheralcoholsareproducedby microbes during fermentation from α -keto acids, by degrading amino acid via "Ehrlich pathway" (Styger et al., 2011).Natural flavours are important quality molecules in a wide range of products like food, cosmetics,beveragesandpharmaceuticals.Nowadays,mostofthemoriginatefromchemicalsynthesisorextractionfrom plants (Carlquist et al., 2014). The 2-phenylethanol (2-PE) aromatic alcohol, with a global production equalingabout 10000 tons per year (Hua and Xu, 2011), is one of the most significant volatile substances.Naturalflavours are important quality molecules in a wide range of products like food, cosmetics, beverages andpharmaceuticals.Nowadays,mostofthemoriginatefromchemicalsynthesisorextractionfromplants(Carlquist et al., 2014). The 2-phenylethanol (2-PE) aromatic alcohol, with a global production equaling about10000tons peryear (Huaand Xu,2011),is oneofthemostsignificantvolatilesubstances.

Key Words: Citrus, Orangepeels, Limonene, 2-phenylethanol, Microbes, *Aspergillusniger*, Fermentation, Citric Acid, Aroma, Alcohol.

Received 15 June, 2022; Revised 28 June, 2022; Accepted 30 June, 2022 © The author(s) 2022.

Published with open access at www.questjournals.org

I. INTRODUCTION:

Orange fruit contain 70% to 90% water concentration. Orange like citrus fruits are excellent source of vitaminC, A and B, fiber (pectin, cellulose, hemicelluloses& lignin.), minerals (likepotassium and calcium) anti-oxidantcompoundslikephenolicandcarotenoids(Arnarson,2019).Citrusprocessingindustrygeneratestoneofwasteannually (salma and Ibrahim, 2018). India Produce 25 lakh tonnes of orange every year. Orange peel is a primary waste in production of orange juice so the waste is accumulated in bulk and make environmentalproblems. (Gotmare and Gade, 2018) The concentrations of total aldehydes increasewith fruit maturity butdeclines with over-maturity.Citric acid is one of the most important organic acids. The citrus peels containcyclicmonoterpenelimonene.Limoneneis themajorcomponentin theoilofcitruspeel.Inorangeoilgreater

than 90% D-limonene are present. The D-limonene, occurring common in nature as the fragrance of orange, uses as a flavoring agent in food manufacturing, in cosmetic, and in pharmaceutical they can hide the unpleasant tastes of drug. Mantzouridou et al. (2015) used *Saccharomyces cerevisiae* for solid-state fermentation of orange peel waste to produce yeast flavor and other valuable chemicals. They obtained maximum volatile aroma esters and total polyphenols, respectively, from fermented orange peel. The large amount of this waste is still dumped every year, which causes both economic and environmental problems such as high transportation cost, lack of dumping site, and accumulation of high organic content material. Therefore, more effective and sustainable alternatives for using orange peel wastes (Wikandari et al., 2015; Grohmann and Baldwin, 1992; Marin et al., 2007). Rose is well known plant for aroma production and has number of industrial uses. Roses have many industrial uses along with its antimicrobial properties. Most of roses are used in cosmetic products. Rose has sweet, pleasurable smell, which is favourable by a lot of people most of the people prefer the smell of rose has most recognised and popular aroma in the world. Essential oil of rose is quite expensive, but has major uses in pharmaceutical and medical. It is also used to cure chronic insomnia (Jane, 2015) Quality of sleep is important during pregnancy and can be improved by changing sleep environment both internally and externally. Rose petals, stem, root, leaves all of them have their own properties and are useful in many ways. Many microorganisms are found from rose plant; they are known as endophytic organisms. Rose plant has been studied with respect to its endophytic fungi characteristics (Catalina et al., 2007). There are many other products that can be produced by rose, such as dye, cosmetics, essential oils, perfumes, jam, aromatic products, and medicines. In many fermented food products, 2-phenyl ethanol and 2-phenyl acetate are responsible for rose like smell. 2-phenylethanol (2-PE) has highest industrial value for its rose-like smell which is mostly used in cosmetic industries. Some microbial transformation processes are also able to produce 2-PE but its yield is limited due to stress of organic solvents on microorganisms. 2-PE can naturally obtain from flowers like rose, but the extraction method is costly. Market price of naturally produced 2-PE is \$1000/kg whereas, chemically production of 2-PE cost \$5/kg, therefore 2-PE bio-production has gotten more attention (Xinyao et al., 2016). There are bundle of microorganisms are present which can produce 2-PE, such as bacteria, Fungi, and yeast (Masuo et al., 2015 and Gathins et al., 2015).

II. PROPOSED METHODOLOGY

CHEMISTRY OF ORANGE PEELS:

Orange Peels are receiving great attention for their high concentration of limonene. This compound is a cyclic monoterpene ($C_{10}H_{16}$), presenting two different optical isomers: Limonene & L-limonene. An interest in the production of enzymes for recovery of oil from the orange peels is very attractive. The methods employ either submerged fermentation or solid-state fermentation (SSF), the latter being more common. In this study, solid-state fermentation of fresh orange peels by *Aspergillus* sp. to extract extra-cellular enzymes and orange peels oil are noted and the effect of methods of orange peel sterilization, importance of selection of the fermentation strain, external nitrogen source, and particle size of the peels are also studied (Labrath & Gaikar, 2020).

CHEMISTRY OF ROSE PETALS:

In 1913, it was obtained the first patent in the United States for a method of producing citric acid by *Aspergillus niger* in sugar solutions (Soccol et al., 2006). 2-Phenylethanol (2-PE) is an important flavor and fragrance compound with a rose-like odour. The natural 2-phenylethanol is mainly extracted from rose petals that involve a high-cost process. Specific strains of yeast like *Saccharomyces cerevisiae*, *Hansenula anomala* are also capable of producing aromatic compounds by bioconversion of 2-phenylalanine into 2-phenylethanol (Stark et al., 2002).

SUBSTRATE:

Sweet oranges and red roses were used as a sample for the isolation of microorganisms, which was collected from the different regions. Yeast was isolated from slightly rotten sweet orange peels which was left in the air tight plastic bag for two days and then were soaked in sterilized distilled water for 3 hours before used as primary sample. Fungi was isolated from rose petals which was left in the open for two days and then soaked in sterile distilled water for another 3 hours. Orange press was obtained from a citrus processing fruit using FMC juice extractors (FMC Corporation, Florida division, Lakeland FL, USA). The liquor was centrifuged at 2100 gm for 10 min and the supernatant was used as the fermentation medium.

CULTURES:

Orange peel and rose petals were cut into small pieces using sterile scalpel and placed into distilled water separately for three hours. After serial dilution of sample, it was then spread onto different agar plates such as GYE agar plate for yeast, Sabouraud's agar plate for fungi. Orange peels a

mple and rose petal sample were spread on GYE agar plate and sabouraud's agar plate respectively. All the plates were incubated for 24-Hours. The citric acid-producing strains of *Aspergillus niger* NRRL567 and NRRL599 were obtained from the CAB International Mycological Institute (Surrey, UK) and the strain NRRL567 from the American Type Culture Collection (Rockville, MD, USA). Each culture was grown on potato dextrose agar slants (Merck) for 5 days. A spore inoculum was prepared by adding 7 ml of sterilized distilled water to each slant and shaking vigorously for 1 min.

III. FERMENTATION MEDIUM

The pH of the OPL medium was adjusted to 3.0-6.2 with concentrated hydrochloric acid prior to sterilization. The medium 100 g was dispensed into 250 ml Erlenmeyer flasks. The flasks were autoclaved at 20 min. Each flask was inoculated with 0.4 ml of the inoculum. Methanol (0, 20, 40, 60 ml) was added and flasks were incubated at 200 rpm. All samples were prepared in triplicate.

ANALYTICAL METHODS:

GC MS assay method for 2-phenylethanol headspace was analyzed using a gas chromatography with Agilent technologies technology with a capillary column HP-5. Citric acid and phenyl ethanol levels in the experiments were determined by studying the kinetic profile of citric acid production using a HPLC and Thin layer chromatography. Sweet oranges and red roses were used as a sample for the isolation of microorganisms. Orange peel sample and rose petals sample were spread on GYE agar plate and sabouraud's agar plate respectively. All the plates were incubated for 24-hour at 37°C. After incubation for 24 hours at 37°C all the plates were observed carefully for their colony characteristics. After identification of microorganisms two different fermentation broths were prepared. Fruit waste fermentation broth with yeast extract in 100 ml water with orange peel powder as raw material was prepared for the fermentation of citric acid and limonene. Floral waste fermentation broth with yeast extract in 100 ml water with fresh rose petals as raw material was prepared for the fermentation of β -phenyl ethanol. Isolated yeast, fungi and cyanobacteria were inoculated in both kind of fermentation broths separately and were incubated at 37°C for five days. Determination of different product was performed after every 24 hours and result were noted. Titration method is used to determine the amount of alcohol in fermentation broth. This is widely used method. Citric acid estimation was carried out using acid-based titration method. The reason we used titrimetric method was because it is simple, accurate, rapid and cost-effective method and can be carried out in any simple laboratory. Oil recovery and estimation is quite difficult method and limonene is a terpene which makes it more difficult to analyses. Bromate titration method is very easy to handle and give accurate results. β -phenyl ethanol can easily vaporize in the air and difficult to keep without a little cold temperature. We used ice-bath to keep it cold during the estimation process. For standard, extract of rose petals was used. estimation and extraction of products in alcohol, phenol, citric acid, limonene. Phytochemical analysis in Carbohydrate test in blue precipitate, Tannins test in greenish blue, Saponins test foam production, Glycoside test red and blue separate layers, Flavonoid test yellowish orange, Alkaloid test in red-brown color.

IV. DETERMINATION

ALCOHOL:

Alcohol is one of the main contents produced during fermentation. Titration method is used to determine the amount of alcohol in fermentation broth. This is widely used method. Alcohol is calculated using alcohol standard curve. The process starts with 1 ml aliquot taking in test tube. 4 mL distilled water was added with 10 ml of 0.2N K₂Cr₂O₇. Tubes were incubated in boiling water-bath for 30 min. After incubation and cooling it down to the room temperature 20% KI (4 ml) was added and mixed thoroughly. Solution was titrated against 0.1N Na₂S₂O₃ till the colour changes to wine red, then few drops of 1% starch was added and colour turned to bluish-green which was again titrated against 0.1N Na₂S₂O₃ till the colour change to light blue. Value of B-E was calculated and plotted in graph (Shinde and Patil, 2016).

PHENOL:

The Folin-Ciocalteu method is described in various pharmacopeia for determination of phenolic compounds. Blue colour is formed during the reaction because of phosphotungstic-phosphomolibdenum complex, where the maximum absorption depends on the concentration of phenolic compound. The Folin-Ciocalteu was used to perform phenol estimation test. For the reference chemical standard, stock solution of gallic acid (20 μ g/ml) prepared in distilled water was used, and for the test sample, incubated fermentation broths were used with undiluted and 1:10 diluted sample. 1 mL of each solution was transferred to separate tubes containing 1 mL Folin-Ciocalteu reagent. All the tubes were kept at room temperature for 3 min and then 10% Na₂CO₃ (10 mL) was added. Tubes

were incubated for 30 min at room temperature. After incubation samples were analyse inUV-Vis spectrophotometer forabsorbance at740nm,waterwasusedas blank.
(Melloetal.,2013)

CITRICACID:

Citric acid estimation was carried out using acid-based titration method.The reason we used titrimetricmethodwasbecauseitissimple,accurate,rapidandcost-effective method and can be carried out in any simple laboratory. Sample was collected from fermentation broth by directly filter it and then centrifuge the sampleat 1000 rpm for 5min. For standard, citric acid was directly extracted out from orange peel, and differentconcentrationswerepreparedandanalyzed.First20mLsamplewastakeninflaskandfewdropsofphenolphthalein indicator was added. Solution was titrated using 0.1N NaOH solution. Final observation was colour change from colorless to pink indicates the presence of citric acid. Final volume of NaOH was noteddown. (Eidetal., 2014)

LIMONENE:

Oilrecoveryandestimationisquitedifficultmethodandlimoneneisaterpenewhichmakesitmoredifficulttoanalyse.Bromatetitrationmethodisveryeasytohandleandgiveaccurateresults.Titrationmethod:
Anemptyglassbottlewasweighedonweighingmachine.5mLfermentationbrothwastakeninbottleandwasweighagain,then15mL2-propanol wasaddedandweighagain, andthesolutiontransferredintoflask. Then10mLmethylorangeindicatorpreparedinHClwas addedandtitratedagainst0.1Npotassiumbromide-bromate solution till the colour changed to yellowish- orange. Standard was performed using extract fromorangepeel.Amountoflimonene wascalculatedusing simpleformulaas follows:
Amountoflimonene=(0.0034)(Bn)(100)(An+En)/(W)(En)Where,
0.0034=0.1NKBrO₃/KBr
Bn=Voloftitrantused(B-Evalue)

En=Weightofemulsion(Eg-Ag)

Eg=Weightofbottle+alcohol+sampleAg=Weightofbottle +alcohol

An= Weight of alcohol (Ag - T)T=weightofbottle

W= Weight of alcohol emulsion (Eg - T)(Scottetal.,1966)

β -PHENYLETHANOL:

β -phenyl ethanolcaneasilyvaporizeintheairanddifficulttokeepwithoutalittlecoldtemperature.Weusedice-bath to keep it cool during the estimation process. For standard, extract of rose petals was used.In thiscolorimetricmethod5mLfermentationbrothwastakenintotheflaskwith10% solutionofKNO₃inH₂SO₄(2mL). Flask was then placed into ice-bath for 30 min and then transferred into water-bath for another 30 min.Letitcooland4mLNH₄OHand9mLH₂Owasadded,mixedwell.Absorbancewasmeasuredat540nm(McFarlane andThompson.,1964)

V. APPLICATION

Traditionally, (+)-limonene is used as a flavoring compound in citrus-flavored products such as soft drinksandcandyandasfragranceingredientin householdcleaningproductsandperfumes(Duetz etal.,2003). Asaflavourandfragranceingredient,limonenehasarelativelyhighpricebecauseofthe quality requirementsin this field. For this application, chirality is important. (+) Limonene (also called R- or d-limonene) has apleasant, orange-like odor whereas the (-) form (also called S- or l-limonene) has a harsher turpentine-likeodor (Friedman and Miller, 1971). Limonene has minor applications in other products. For example, it is usedasinsecticide(Ciriminnaetal.,2014)andisbeinginvestigatedformedicalapplicationsduetoitsantimicrobialandanti-cancer properties(Inouyeetal.,2001;Milleretal., 2010)

Citricacidmainlyusedinfoodindustry,pharmaceuticals,chemicalindustry,cosmetics,printings,foodpreservative, electro pickling, copper plating, beverage & others. Some specific applications are given below(Soccol,2003; Pandeyetal.,2001; Vandenberghe,1999;Grewal,1995).

1. Citricacidmonohydrateiswidelyusedasorganicacid&pHcontrolagent,flavouringandpreservativeinfoodproductionlikeascandy,cookies,biscuits,jams,jellies,snacks,instantfoodsandsauces.
2. It is used as acidity regulator and antioxidant in beverage such as alcoholic beverage, carbonated soft drink,syrups,juice drinks, tea&coffee, ice-cream,sports&energydrink.
3. Itcanbeusedinthrombininhibitorandfungicideinpharmaceutical.

Anti-Infective Agents, 2-phenylethanol on humans and other animals that destroy harmful microorganisms or inhibit their activity. They are distinguished from disinfectants, which are used on inanimate objects. Use as Preservatives, in pharmaceutical preparation to protect them from chemical change or microbial action. They include anti-bacterial agents and antioxidants (Corre, 1990).

VI. CONCLUSION

In 1913, it was obtained the first patent in the United States for a method of producing citric acid by *Aspergillus niger* in sugar solutions (Soccol et al., 2006). Orange peel was employed in this work as raw material for the production of citric acid (CA) by solid-state fermentation (SSF) of *Aspergillus niger* CECT-2090 (ATCC9142, NRRL599) in Erlenmeyer flasks (Tornado et al., 2011). Microbial production of citric acid and limonene using orange peel powder and β -phenyl ethanol using rose petal shave shown great amount of production by yeast *Saccharomyces* sp., fungi *Aspergillus* sp. Microbial production of citric acid, limonene, 2-phenylethanol has been carried out successfully using various method, still confirm analysis using GC-MS or FTIR or TLC is necessary. Even though, it is quite difficult to work the microorganisms we used gave quite the satisfactory results. Microbial production of citric acid, limonene, 2-phenylethanol has been carried out successfully using various method. In future aspect genetic analysis of microorganisms, anti-fungal & anti-bacterial. The medium used throughout this study contained total sugars as glucose: fructose: sucrose ratio of 1.0; 0.8; 0.7, respectively.

REFERENCES:

- [1]. Rivas, B., Torrado, A., Torre, P., Converti, A., & Domínguez, J. M. (2008). Submerged citric acid fermentation on orange peel autohydrolysate. *Journal of Agricultural and Food Chemistry*, 56(7), 2380-2387.
- [2]. Torrado, A. M., Cortés, S., Salgado, J. M., Max, B., Rodríguez, N., Bibbins, B. P., ... & Domínguez, J. M. (2011). Citric acid production from orange peel wastes by solid-state fermentation. *Brazilian Journal of Microbiology*, 42(1), 394-409.
- [3]. Chreptowicz, K., Wielechowska, M., Główczyk-Zubek, J., Rybak, E., & Mierzejewska, J. (2016). Production of natural 2-phenylethanol: From biotransformation to purified product. *Food and Bioproducts Processing*, 100, 275-281.
- [4]. Etschmann, M. M. W., Sell, D., & Schrader, J. (2003). Screening of yeasts for the production of the aroma compound 2-phenylethanol in molasses-based medium. *Biotechnology Letters*, 25(7), 531-536.
- [5]. Etschmann, M., Bluemke, W., Sell, D., & Schrader, J. (2002). Biotechnological production of 2-phenylethanol. *Applied Microbiology and Biotechnology*, 59(1), 1-8.
- [6]. Soccol, C. R., de Souza Vandenberghe, L. P., Medeiros, A. B. P., Karp, S. G., Buckeridge, M., Ramos, L. P., ... & Torres, F. A. G. (2010). Bioethanol from lignocelluloses: status and perspectives in Brazil. *Bioresource Technology*, 101(13), 4820-4825.
- [7]. Torrado, A. M., Cortés, S., Salgado, J. M., Max, B., Rodríguez, N., Bibbins, B. P., ... & Domínguez, J. M. (2011). Citric acid production from orange peel wastes by solid-state fermentation. *Brazilian Journal of Microbiology*, 42(1), 394-409.
- [8]. Etschmann, M. M. W., & Schrader, J. (2006). An aqueous-organic two-phase bioprocess for efficient production of the natural aromachemicals 2-phenylethanol and 2-phenylethyl acetate with yeast. *Applied Microbiology and Biotechnology*, 71(4), 440-443.
- [9]. Labrath, Y. P., & Gaikar, V. G. (2020). Solid State Fermentation of Orange Peels for Production of Cellulase, Pectinase and Recovery of Orange Oil using *Aspergillus* Species NCIM1432.
- [10]. Correa, C.R., & Kruse, A. (2018). Super critical water gasification of biomass for hydrogen production—Review. *The Journal of Supercritical Fluids*, 133, 573-590.
- [11]. Saerens SMG, Verstrepen KJ, Van Laere SDM, Voet ARD, Van Dijck P, Delvaux FR, Thevelein JM (2006) The 80 *Saccharomyces cerevisiae* EHT1 and EEB1 genes encode novel enzymes with medium-chain fatty acid ethyl ester synthesis and hydrolysis capacity. *The Journal of Biological Chemistry*
- [12]. Styger G, Prior B, Bauer FF (2011b) Wine flavor and aroma. *Journal of Industrial Microbiology and Biotechnology*. Vol 38, PP: 1145-1159. ISSN: 1367-5435.
- [13]. Eden A, Van Nedervelde L, Drukker M, Benvenisty N, Debourg A (2001) Involvement of branched-chain amino acid aminotransferases in the production of fusel alcohols during fermentation in yeast. *Applied Microbiology and Biotechnology*(55),PP: 296-300. ISSN: 1432-0614
- [14]. Styger G, Jacobson D, Bauer FF (2011a) Identifying genes that impact on aroma profiles produced by *Saccharomyces cerevisiae* and the production of higher alcohols. *Applied Microbiology and Biotechnology*(91), PP: 713-730. ISSN: 1432-0614.
- [15]. JaneBucklePhD, RN, in *Clinical Aromatherapy* (Third Edition), 2015. ISBN: 9780702064869.
- [16]. Xinyao Lu, Yuqin Wang, Hong Zong, Hao Ji, Bin Zhuge, and Zhuoli Dong (2016). Bioconversion of phenylalanine to 2-phenylethanol by the novel stress-tolerant yeast *Candida* glycerinogenes WL2002-5. *Bioengineered*. Vol 7(6): 418-423. ISSN: 2165-5987.
- [17]. Masuo S, Osada L, Zhou S, Fujita T, Takaya N. (2015). *Aspergillus oryzae* pathways that convert phenylalanine into the flavour volatile 2-phenylethanol. *Fungal Genetics and Biology*. Vol 77, PP: 22-30. ISSN: 1087-1845. <http://dx.doi.org/10.1016/j.fgb.2015.03.002>
- [18]. Gethins L, Guneser O, Demirkol A, Rea MC, Stanton C, Ross R P, Influence of carbon and nitrogen source on production of volatile fragrance and flavour metabolites by the yeast *Kluyveromyces marxianus*, *Yeast* (2015); Vol 32: PP: 67-76; ISSN: 1097-0061.
- [19]. Wikandari R., Nguyen H., Millati R., Niklasson C., & Taherzadeh M. J. (2015). Improvement of biogas production from orange peel waste by leaching of limonene. *BioMed research international*, Vol 2015. ISSN: 2314-6141 <https://doi.org/10.1155/2015/494182>
- [20]. K. Grohmann and E. A. Baldwin (1992). Hydrolysis of orange peel with pectinase and cellulase enzymes. *Biotechnology Letters*, vol. 14, no. 12, PP: 1169-1174, ISSN: 1573-6776. <https://doi.org/10.1007/BF01027023>
- [21]. F. R. Marin, C. Soler-Rivas, O. Benavente-García, J. Castillo, and J. A. Pérez-Alvarez (2007). By-products from different citrus processes as a source of customized functional fibres. *Food Chemistry*, vol. 100, no. 2, PP: 736-741,

- ISSN:0308-8146.
- [22]. Battista F., Remelli G., Zanzoni S., & Bolzonella D. (2020). Valorization of residual orange peels: Limonenerecovery, volatilefattyacids, andbiogasproduction. *ACSSustainableChemistry&Engineering*, 8(17), PP:6834-6843. ISSN:2168-0485https://doi.org/10.1021/acssuschemeng.0c01735
- [23]. Labrath Y. P., & Gaikar V. G. (2020). Solid State Fermentation of Orange Peels for Production of Cellulase, Pectinase and Recovery of Orange Oil using Aspergillus Species NCIM1432. doi: 10.21203/rs.3.rs-123470/v1
- [24]. Rivas B.; Torrado A.; Torre P.; Converti A.; Domínguez J.M. (2008). Submerged citric acid fermentation on orange peel autohydrolysate. *Journal of Agriculture and food Chemistry*. 56, PP:2380–2387. ISSN:1520-5118.
- [25]. Soccol C.R.; Vandenberghe L.P.S.; Rodrigues C.; Pandey A. (2006). New Perspectives for Citric Acid Production and Application of Food Technology and Biotechnology. 44, PP:141–149. ISSN:1330-9862.
- [26]. Kapoor K.K.; Chaudhary K.; Tauro P. (1982). *Prescott and Dunn's Industrial Microbiology*, 4th edn. G. Reed (Ed), AVI Publishing Co, Westport, CT. https://doi.org/10.1023/A:1008821413239
- [27]. Torrado A. M., Cortés S., Salgado J. M., Max B., Rodríguez N., Bibbins B. P., ... & Domínguez J. M. (2011). Citric acid production from orange peel wastes by solid-state fermentation. *Brazilian Journal of Microbiology*, Vol 42, PP:394-409. ISSN:1517-8382.
- [28]. Roccia A., Hibrand-Saint Oyant L., Cavel E., Caillard J. C., Machenaud J., Thouroude T. & Baudino S. (2019). Biosynthesis of 2-phenylethanol in rose petals is linked to the expression of one allele of RhPAAS. *Plant Physiology*, Vol 179(3), PP:1064-1079. ISSN: 0032-0889
- [29]. Carlquist M., Gibson B., Yuceer Y.K., Paraskevopoulou A., Sandell M., Angelov A.I. (2014). Process engineering for bioflavour production with metabolically active yeasts minireview. *Yeast*. 32, PP:123-143. ISSN:1097-0061. https://doi.org/10.1002/yea.3058
- [30]. Kovacheva N., Rusanov K., Atanassov I. (2010). Industrial Cultivation of Oil Bearing Rose and Rose Oil Production in Bulgaria during 21st Century. *Directions and Challenges in Biotechnology and Biotechnological Equipment*. Vol 24, PP: 1793-1798. ISSN: 1314-3530.
- [31]. Cui Z., Yang X., Shen Q., Wang K., Zhu T., (2011) Optimization of biotransformation conditions for production of 2-phenylethanol by a *Saccharomyces cerevisiae* CWY132 mutant. *Nat. Prod. Res.* 25, PP: 754-759. ISSN:2673-1665.
- [32]. Huad., Xu P. (2011). Recent advances in biotechnological production of 2-phenylethanol. *Biotechnology Advances*. Vol 29, PP: 654-660. https://doi.org/10.1159/000455169
- [33]. Garavaglia J., Flores S., Pizzolato T., Peralba M., Ayub M. (2007). Bioconversion of L-phenylalanine into 2-phenylethanol by *Kluyveromyces marxianus* using grape must cultures. *World Journal of Microbiology and Biotechnology*. Vol 23, PP: 1273-1279. ISSN: 1573-0972.
- [34]. Huang C.J., Lee S.L., Chou C.C. (2000). Production and molar yield of 2-Phenylethanol by *Pichia fermenter* L-5 as affected by some medium components. *Journal of Bioscience and Bioengineering*. Vol 90(2), PP:142-147, ISSN: 1389-1723.
- [35]. Yin S., Zhou H., Xiao X., Lang T., Liang J., Wang C., (2015.) Improving 2-phenylethanol production via Ehrlich Pathway using genetic engineered *Saccharomyces cerevisiae* strains. *Current Microbiology*. 70, PP: 762-767. ISSN:1432-0991. https://doi.org/10.1159/000455169
- [36]. Kim B., Cho B.R., Hahn J.S. (2014a). Metabolic engineering of *Saccharomyces cerevisiae* for the production of 2 phenylethanol via Ehrlich pathway. *Biotechnology and Bioengineering*. Vol 111, PP:115-124. ISSN:0006-3592.
- [37]. Stark D., Munch T., Sonnleitner B., et al. Extractive bioconversion of 2-phenylethanol from L-phenylalanine by *Saccharomyces cerevisiae*. *Biotechnol Prog*. (2002); Vol 18(3) PP: 514–523. ISSN:8756-7938. https://doi.org/10.1021/bp020006n
- [38]. Zanetti M., Ternus ZR., Dalcanton F., de Mello MMJ., de Oliveira D., et al. (2015) Microbiological Characterization of Pure Geraniol and Comparison with Bactericidal Activity of the Cinnamic Acid in Gram-Positive and Gram- Negative Bacteria. *Journal of Microbiology, Biochemistry and Technology*. 7:4P: 186-193. https://doi.org/10.3390/ijms22041717
- [39]. Li R., Wang K., Wang D., Xu L., Shi Y., Dai Z., & Zhang X. (2021). Production of plant volatile terpenoids (rose oil) by yeast cell factories. *Green Chemistry*. https://doi.org/10.3390/molecules1912 19292
- [40]. National Center for Biotechnology Information (2021). PubChem Compound Summary for CID 22311, Limonene. Retrieved July 26, 2021 from https://pubchem.ncbi.nlm.nih.gov/compound/Limonene.
- [41]. Koolaji N., Shammugasamy B., Schindeler A., Dong Q., Dehghani F. & Valtchev P. (2020). Citrus peel flavonoids as potential cancer prevention agents. *Current developments in nutrition*, Vol 4(5), PP:25. ISSN:2475-2991. https://doi.org/10.1093/cdn/nzaa025
- [42]. National Center for Biotechnology Information (2021). PubChem Compound Summary for CID 311, Citric acid. Retrieved July 26, 2021 from https://pubchem.ncbi.nlm.nih.gov/compound/Citric-acid.
- [43]. National Center for Biotechnology Information (2021). PubChem Compound Summary for CID 6054, 2- Phenylethanol. Retrieved July 26, 2021 from https://pubchem.ncbi.nlm.nih.gov/compound/2-Phenylethanol.
- [44]. Boluda-Aguilar M., Lopez-Gomez A. (2013). Production of bioethanol by fermentation of lemon (*Citrus limon* L.) peel wastes pretreated with steam explosion. *Industrial Crops and Products*. Vol 41, PP:188–197. ISSN:0926-
- [45]. L.P.S. Vandenberghe, C.R. Soccol, A. Pandey, J.M. Lebeault (1999). Review: Microbial production of citric acid. *Brazilian Archive of Biology and Technology*. Vol 42, PP:263–276. https://doi.org/10.1590/S1516-89131999000300001.
- [46]. W.A. Duetz, A.H. Fjällman, S. Ren, C. Jourdat, B. Witholt. (2001). Biotransformation of d-Limonene to (+)-trans-Carveol by Toluene-Grown *Rhodococcus opacus* PWD4 Cells. *Applied and environmental microbiology*. 67(6): PP:2829-2832. ISSN: 1098-5336.
- [47]. Nautical O.H., Tiwari K.K. (2012). Supercritical Carbon Dioxide Extraction of Indian Orange Peel Oil and Hydro Distillation Comparison on Their Compositions. *Science and Technology*. 2(3) PP:27-31. DOI: 10.5923/j.scit.20120203.01.
- [48]. Scalliet G., Journot N., Jullien Fetal. (2002). Biosynthesis of the major scented components 3,5-dimethoxytoluene and 1,3,5-trimethoxybenzene by novel rose O-methyltransferases. *FEBS Letters*. Vol 523, PP:113–118. ISSN:0014-5793.
- [49]. Rayne S., Forest K. (2016). Carboxylic acid ester hydrolysis rate constants for food and beverage aroma compounds. *Flavour and Fragrance Journal*. Vol 31, PP:https://doi.org/10.1002/ffj.3327.
- [50]. Jakobsen HB., Olsen CE. Influence of climatic factors on emission of flower volatiles in situ. *Planta*. (1994);192:365–371. https://doi.org/10.1007/BF00198572.
- [51]. V.A. Shinde and R.B. Patil (2016). Production of Ethanol by *Saccharomyces cerevisiae* Using Orange Peels and Banana Peels. *International Journal of Current Microbiology and Applied Sciences*, Vol:5 PP:280-284, ISSN:2319-7706107.
- [52]. Scott, W. Clifford and M.K. Veldhuis (1966). Rapid estimation of recoverable oil by bromate titration. *Journal of Association of Official Analytical Chemists*, ISSN:0004-5756.
- [53]. Mohammed Shaibu Auwal, Sanni Saka, Ismail Alhaji Mairiga, Kyari Abba Sanda, Abdullahi Shuaibu and Amina Ibrahim (2014). Preliminary phytochemical and elemental analysis of aqueous and fractionated pod extracts of *Acacia*

- nilotica (Thornmimosa), Veterinary research forum,5(2): 95–100, PMID: 25568701.
- [54]. W.D.McFarlane and K.D.Thompson(1964).Colorimetric methods for determination of aromatic alcohols Journal of the Institute of Brewing, 70: 497-502. in beer.
- [55]. Joseph K. Adu, Cedric D. K. Amengor, Naomi Kabiri, Emmanuel Orman, Stella Abla Gameli Patamia, and Bernice Korkor Okrah(2019). Validation of a Simple and Robust Liebermann–Burchard Colorimetric Method for the Assay of Cholesterol in Selected Milk Products in Ghana, International Journal of Food Science, Volume 2019, Article ID: 9045938.
- [56]. Christy Gnana Theebap, and Sasi Kumar R.(2015). Phytochemical examination, antioxidant potential and invitro antibacterial studies of crud extracts of Parthenium hysterophorus Linn. Leaves, Journal of Chemical and Pharmaceutical Research, 7(4):219-225, ISSN:0975-7384.
- [57]. Xinyao Lu, Yuqin Wang, Hong Zong, Hao Ji, Bin Zhu Ge, and Zhuoli Dong, Bioconversion of -phenylalanine to 2-phenylethanol by the novel stress-tolerant yeast *Candida glycerinogenes* WL2002-5 Bioengineered. 2016;7(6):418–423.
- [58]. Joseph K. Adu, Cedric D. K. Amengor, Naomi Kabiri, Emmanuel Orman, Stella Abla Gameli Patamia, and Bernice Korkor Okrah(2019). Validation of a Simple and Robust Liebermann–Burchard Colorimetric Method for the Assay of Cholesterol in Selected Milk Products in Ghana, International Journal of Food Science, Volume 2019, Article ID: 9045938.
- [59]. Braddock RJ, Cadwallader KR 1992 Citrus by-products manufacture for food use. Food Technol 46(2):105-110.
- [60]. Battista F., Remelli G., Zanzoni S., & Bolzonella D. (2020). Valorization of residual orange peels: Recovery, volatile fatty acids, and biogas production. ACS Sustainable Chemistry & Engineering, 8(17), PP:6834-6843. ISSN:2168-0485 <https://doi.org/10.1021/acssuschemeng.0c01735>.